



**Pest risk assessment of the American Serpentine Leafminer
(*Liriomyza trifolii*) in Norway**

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Liriomyza trifolii (Burgess) in Norway

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SUMMARY

Liriomyza trifolii Burgess is a pest species that originates from Central and North America, but since the 1970-ies it has spread with plants to many parts of the world. In the tropics, subtropics and warmer parts of the temperate zone it has been established in the field, while in a colder climate it can develop as a pest only in greenhouses. The pest has a wide host plant range. In Europe the pest has been reported in most countries, Norway included; predominantly on vegetables imported from Asia.

The pest risk assessment was initiated by the Norwegian Scientific Committee for Food Safety (Vitenskapskomiteen for mattrygghet, VKM) Panel on Plant Health.

The VKM Panel on Plant Health gives the following main conclusions of the risk assessment: 1) *L. trifolii* has been spread to Norwegian greenhouses on several occasions, but each time it has been eradicated. It is not present in Norway today. 2) The overall probability of entry of *L. trifolii* into Norway and the overall probability of establishment in greenhouses of *L. trifolii* in Norway are both rated as high with low levels of uncertainty. 3) In the absence of statutory control the probability for *L. trifolii* to be spread quickly in greenhouses in the PRA area by trade of host plants is rated as high. The uncertainty of this assessment is low. 4) *L. trifolii* can be established in the field around infested greenhouses during the summer, but it can not overwinter in the field in Norway. The level of uncertainty of this assessment is low. 5) The part of the PRA area where presence of *L. trifolii* might result in economically important losses (the endangered area) in greenhouses is assessed to be all of Norway. 6) *L. trifolii* is likely to have moderate economic impact in the greenhouses in the PRA area with current phytosanitary measures. Without any such regulations *L. trifolii* is likely to have major economic impact on the greenhouse industry of the PRA area. The levels of uncertainty of these assessments are low. 7) The non-commercial and environmental consequences to natural environments in the PRA area are likely to be low. The level of uncertainty of this assessment is low.

CONTRIBUTORS

Persons working for VKM, either as appointed members of the Committee or as *ad hoc* experts, do this by virtue of their scientific expertise, not as representatives for their employers. The Civil Services Act instructions on legal competence apply for all work prepared by VKM.

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1. BACKGROUND

Among the three polyphagous *Liriomyza*-species that are quarantine pests in Norway (*L. huidobrensis*, *L. sativae* and *L. trifolii*), *L. trifolii* has higher temperature preferences than *L. huidobrensis*, and almost as high temperature preferences as *L. sativae* (Tokumaru & Abe 2003). It has been established in many tropical, subtropical and warmer parts of the temperate areas all over the world (except Australia). In addition there are regular outbreaks in greenhouses in most parts of the world. The larvae of *L. trifolii* are highly polyphagous, being able to develop inside the leaves of plants in many plant families, including cultivated plants like many vegetables, ornamentals and cotton. Under favorable climatic conditions generations follow in quick succession, and serious damage has been reported in many agricultural and ornamental crops.

There have been some changes in the global distribution of *L. trifolii* (Appendix 1) since the last PRA was made for the pest in Norway (Sæthre 1996). In Europe the pest has been reported in many countries, predominantly on imported vegetables from Asia (Appendix 2). Due to different levels of investigation and policies in different countries, the current distribution maps of the species in Europe and the rest of the world (EPPO 2006) are not well documented, and the information must be used with caution.

In Norway there have been several incidents of the pest in greenhouses since 1978. In all cases the populations have been eradicated, but the incidents have made the Norwegian Food Safety Authority on the alert concerning the species.

The report from the *ad hoc* group has been initiated, evaluated and approved by VKM Panel on Plant Health. The pest risk assessment was adopted by VKM's Plant Health Panel on a meeting December 10th 2009.

Be aware that the current document is a pest risk assessment, and not a Pest Risk Analysis (PRA). A PRA consists of both a risk assessment and a risk management part. VKM performs purely the risk assessment, whereas the Norwegian Food Safety Authority is responsible for the risk management. However, since this pest risk assessment is part of a PRA process, the current document refers to the PRA term in several contexts, like the identification of the PRA area and referrals to former PRAs. This is in accordance with the international standard ISPM No. 11 (FAO 2004).

2. INITIATION

2.1. Initiation points

2.1.1. PRA initiated by the identification of a pest

Initiated by the Norwegian Scientific Committee for Food Safety, a previous Norwegian PRA is being re-evaluated. The pest has been established in many countries all over the world, and it has been spread to Norway several times. Also, the taxonomy of the pest has recently been investigated. Consequently, the timing of the PRA initiation is due to repeated entries into Norwegian greenhouses in recent years, and to new knowledge about the pest.

2.2. Identification of PRA area

The PRA area is Norway.

2.3. Information

Information sources utilised for this pest risk assessment are published material available in international scientific journals, books and reports, as well as personal communications with persons involved in the area, geographical data, unpublished results, and information from the Norwegian Food Safety Authority that have been made available to the risk assessors. Where these information sources have been used, this is indicated in the text by references enclosed in brackets.

The current pest risk assessment is made according to the international standard ISPM No. 11 (FAO 2004).

2.3.1. Previous PRAs

Commissioned by the former Norwegian Agricultural Inspection Service, the former Norwegian Crop Research Institute (Planteforsk) in 1996 did a PRA on *Liriomyza trifolii* (Sæthre 1996). The PRA referred to the first observation of the pest in Norwegian greenhouses in 1978, but fail to mention the incidents in 1980 and 1982. The biology of the species was given, and available control measures and the potential economic importance were evaluated.

The PRA was followed up by an investigation of possible *Liriomyza* species being present in greenhouses and the field in Norway in 1996 (Sæthre 1997) and 2003-2005 (Johansen *et al.* 2004, 2006).

Important information is also found in the two EPPO documents “EPPO Data Sheet on Quarantine Pests. *Liriomyza trifolii*” (EPPO 1997) and “EPPO Diagnostic. *Liriomyza* spp.” (EPPO 2005).

2.4. Conclusion of initiation

The pest of concern is the dipterous pest *Liriomyza trifolii*. The work was initiated by the Norwegian Scientific Committee for Food Safety, and the initiation point for the pest risk assessment is the re-evaluation of a previous PRA for Norway. The PRA area is Norway.

3. PEST RISK ASSESSMENT

3.1 Pest categorization

3.1.1. Identity of pest

3.1.1.1 Scientific name

Liriomyza trifolii Burgess, 1880

3.1.1.2 Synonyms

Liriomyza alliovora Frick, 1955

Liriomyza trifolii de Meijere, 1925

Oscinis trifolii Burgess, 1880

3.1.1.3 Common names

American Serpentine Leafminer

Chrysanthemum Leafminer

3.1.1.4 Taxonomic position

Class: Insecta; Order: Diptera; Family: Agromyzidae; genus: *Liriomyza*.

While *L. trifolii* populations all over the world today are treated as one species, Reitz & Trumble (2002) suggested the existence of cryptic species. Scheffer & Lewis (2006) concluded that the presence of 2-3 different mitochondrial clades in the species is suggestive of cryptic species. However, most of the diversity appeared in the native areas in the Americas, while all the invasive populations belonged to the same phylogenetic clade.

L. trifolii is taxonomically also very closely related to *L. sativae*, another invasive species. This sometimes poses a problem in identification at border controls, especially if only female specimens are found. Shiao (2004) gives valuable information on how to separate the two species morphologically.

As a conclusion, due to the difficult taxonomy of the species and several very closely related species, all information concerning *L. trifolii* has to be evaluated with caution.

3.1.2 Presence or absence in PRA area

L. trifolii has never with certainty been discovered at the Norwegian border. However, flies which in some encounters have only been identified to *Liriomyza* sp. may belong to this species (Table 1). It has been intercepted at least four times in trade of greenhouse plants in the PRA area during 1978-2009 (Table 2). After all interceptions into greenhouses in Norway the species has been eradicated (Sæthre 1997). *L. trifolii* has never been found outdoors in Norway.

Table 1. Imports to Norway the last five years stopped due to records of *Liriomyza* spp. by import control (Norwegian Food Safety Authority).

Year	Pest species	Plant species	Country of origin
2004 – 2007: no records			
2008	<i>L. sp.</i>	<i>Exacum sp.</i>	Denmark
	<i>L. huidobrensis</i>	<i>Exacum sp.</i>	Denmark
	<i>L. huidobrensis</i>	<i>Exacum sp.</i>	Denmark
	<i>L. sp.</i>	<i>Solidago sp.</i>	Zimbabwe
	<i>L. sp.</i>	<i>Verbena sp.</i>	Netherlands
2009: no records			

Table 2. Incidents of *Liriomyza trifolii* into Norwegian greenhouses and garden centres (Norwegian Institute for Agricultural and Environmental Research)

Year	Number of incidents	Number of infested greenhouses and garden centres	Host plant	Country of origin
1978	1	1	<i>Gerbera sp.</i>	Netherlands
1980	2	2	<i>Chrysanthemum sp.</i> and <i>Gerbera sp.</i>	Canary Islands and Netherlands
1982	1	1	<i>Chrysanthemum sp.</i>	Canary Islands
2001	1	1	<i>Gerbera sp.</i>	Netherlands
2001*	1	1	<i>Spinacia oleracea</i>	Sri Lanka

**L. trifolii* or *L. sativae*

3.1.3 Regulatory status

In Norway *L. trifolii* is currently treated as a quarantine pest.

3.1.4 Potential for establishment and spread in PRA area

According to EPPO reports on notifications of non-compliance for *L. trifolii* for the years 2002-2009 (EPPO Reporting Service 2002 – September 2009), it is obvious that there is a high probability that plants containing *L. trifolii* now and then is sought imported into Norway. The occurrence of *L. trifolii* is most common in vegetables from Asia (Appendix 2). Due to the availability of relevant host species and suitable climatic conditions, there is a potential for establishment and spread of *L. trifolii* all year round in greenhouses in the PRA

area. Outdoors in the field it would only survive during the summer. Tokumaru & Abe (2003) showed that the temperature preferences of *L. trifolii* are quite similar to or slightly lower than *L. sativae*. Chen & Kang (2005) suggested a northern overwintering range limit under natural conditions in China for *L. sativae* to be the -2 °C isotherm of the minimum mean temperature in January. In that case, data from Aune (1993) and three meteorological stations in Norway (Table 3) should indicate that *L. trifolii* would be able to overwinter outdoors in the warmest coastal areas of Southern Norway. However, the northern latitude Chen & Kang (2005) suggest for *L. sativae* in China is 34° N, which correspond to south of Crete and Cyprus in Europe and Southern Syria in Asia, far south of Norway. This large difference between China and Europe can be explained by the more Atlantic climate in Europe compared to the continental climate in China and the more Atlantic climate in Europe. We conclude that it is very unlikely that *L. trifolii* would survive during winter even in the mildest parts of Norway.

The mean temperature along the coast of Southern Norway in May – August is around 15 °C, as shown by the mean temperature for three meteorological stations during 1995-2009 (Ås near Oslo in South-Eastern Norway, Særheim near Stavanger in South-Western Norway and Kvithamar near Trondheim in Middle Norway) in Table 3. During 3 months (90 days) at 15 °C, *L. trifolii* should be able to go through slightly less than two generations (Lanzoni *et al.* 2002, Sakamaki *et al.* 2003, Tokumaru & Abe 2003).

Table 3. Monthly mean temperatures (°C) for the years 1995 – 2009 at three sites in coastal Southern Norway (Landbruksmeteorologisk tjeneste (LMT), Bioforsk).

	Jan	April	May	June	July	Aug	Sept	Oct	June-Aug
Særheim	2.4	6.4	9.4	12.3	14.7	15.3	12.5	8.7	14.1
Ås	-2.8	5.0	10.0	14.0	16.1	15.7	11.3	6.1	15.3
Kvithamar	-0.8	5.2	9.1	12.7	15.1	14.8	10.9	6.3	14.2

In conclusion, *L. trifolii* could be spread in greenhouses in the PRA area. The pest could also spread locally in the field around infested areas during the summer and develop into large populations, but the species will not survive the winter in the field.

3.1.5 Potential for economic consequences in PRA area

Yield losses of the three New World *Liriomyza* spp. (*L. huidobrensis*, *L. sativae* and *L. trifolii*) can be significant, and the three species are regarded as serious pests of numerous ornamental and agricultural plants (Parrella 1987, Murphy & La Salle 1999).

In California alone it was estimated that the chrysanthemum industry lost approximately 93 million dollars to *L. trifolii* from 1981 to 1985 (Parrella 1987). For the cut flower industry both the biological and aesthetical damage to plants are important. Aesthetical injury levels have been proposed for floriculture crops including insect pest as *L. trifolii* (Parrella & Jones 1987).

3.1.6 Conclusion of pest categorization

L. trifolii is present in the PRA area only after occasional entries. So far, the pest has been eradicated after every entry.

Due to the availability of hosts and a suitable climate, there is a potential for establishment and spread of *L. trifolii* in greenhouses in the PRA area. All evidence indicates that the species is able to exist and multiply in the field in the summer, but can not survive the winter.

The pest could cause significant loss or damage to plants in greenhouses in the PRA area. Thus, the current pest risk assessment is continued.

3.2. Assessment of the probability of introduction and spread

3.2.1 Probability of entry of the pest

3.2.1.1 Identification of pathways

Pathway A. Import of host plants with eggs, larvae or pupae

L. trifolii might enter the PRA area by import of host plants originating from infested areas. This is shown by the previous history of the pest, especially the high number of infestations detected in vegetables at European borders (Appendix 2). Import of flowers and vegetables are rated as the most likely pathways for imports of *L. trifolii* into the PRA area.

Adults of *L. trifolii* copulate on the host plants, and the females make so-called pinholes by inserting their ovipositor into the leaves to feed on the plant fluids that run from the wounds. Later they lay eggs inside the leaf in some of the pinholes. Larvae hatch from the eggs and create a so-called mine by eating tissue inside the leaf. When fully grown, the larvae leave the mine and pupate either on the outside of the leaf or drop to the ground before they pupate. The next generation of flies emerges from the pupae. Thus, the plant host species offers *L. trifolii* all it needs concerning environment and development. A small infestation can be difficult to discover, since it often can consist only of pinholes, eggs and possibly some larvae in small mines. Also, sometimes the mines are easy to spot from only one side of the leaf, and can easily be overlooked.

The full range of natural host species to date is reported in Appendix 3.

The global distribution of *L. trifolii* is shown in Appendix 1.

Pathway B. Import of soil/growing media with pupae

L. trifolii might be imported into the PRA area with soil/growing media originating from infested areas. *L. trifolii* has the potential to contaminate soil and growing medium as pupae, and the pest has a potential to survive significant periods of time in potting media. The developmental time for pupae depends on the temperature, and is reported as 9.2-8.8 days at 25 °C and 30.2-33.2 days at 15 °C (Lanzoni *et al.* 2002, Sakamaki *et al.* 2003).

Pathway C. Natural spread of adult flies from other European countries by air.

L. trifolii might enter the PRA area by natural spread of adult flies by air from infested areas in other European countries. Wind-borne migration has been shown to exist in many insect taxa, including Diptera species (Gatehouse 1997).

3.2.1.2 Probability of the pest being associated with the pathway at origin

The ratings of probabilities and uncertainties for *L. trifolii* being associated with the pathways at origin are given for each pathway in Table 4. The probabilities varies according to factors like

- prevalence of the pest in the source area
- occurrence of the pest in a life-stage that would be associated with commodities, containers, or conveyances
- volume and frequency of movement along the pathway
- seasonal timing
- pest management
- cultural and commercial procedures applied at the place of origin

Table 4. Estimates of the probability of *L. trifolii* being associated with each pathway at origin in relation to geographical source. The probability of the pest is ranked according to the following scheme: Very unlikely; Unlikely; Moderately likely; Likely; Very likely. Uncertainty for each estimate is given in brackets, and is ranked according to the following scheme: Low; Medium; High.

Pathway	Europe (EU/Switzerland)	USA and Canada	South and Central America	Africa	Asia
A Import of host plants with eggs, larvae, pupae or adult flies	Moderately likely (low uncertainty)	Unlikely (low uncertainty)	Unlikely (medium uncertainty)	Unlikely (medium uncertainty)	Moderately likely (medium uncertainty)
B Import of soil/growing media with pupae	Very unlikely (low uncertainty)	Very unlikely (low uncertainty)	Very unlikely (low uncertainty)	Very unlikely (low uncertainty)	Very unlikely (low uncertainty)
C Natural spread of adult flies by air	Very unlikely (medium uncertainty)	Very unlikely (low uncertainty)	Very unlikely (low uncertainty)	Very unlikely (low uncertainty)	Very unlikely (low uncertainty)

Pathway A. Import of host plants with eggs, larvae or pupae

Generally this is considered as the most probable pathway of entry of *L. huidobrensis* into the PRA area. Due to high import of host plants to Norway from Europe and Asia, the probability of imports from these continents is rated as higher than imports from the rest of the world.

Since the first discovery of *L. trifolii* in Norway in 1978, the pest has never with certainty been stopped at the border (Table 1), but at least four times it has been established for short periods of time in the PRA area greenhouses before it has been eradicated (Table 2).

Pathway B. Import of soil/growing media with pupae

The import of soil and organic growing media into the PRA area is prohibited from countries outside Europe (Landbruks- og matdepartementet 2000). Import of growing medium (except *Sphagnum*) from European countries need to be followed by a Phytosanitary Certificate.

Therefore, the probability for the pest being associated with this pathway at origin is considered as very unlikely from all parts of the world.

Pathway C. Natural spread of adult flies from other European countries by air.

Natural spread of *L. trifolii* by aerial dissemination of adult flies is possible, as strong winds could potentially move *L. trifolii* over great distances from other European countries like Sweden, Denmark, Germany and Poland into the PRA area. Such weather events occur sometimes when there are strong southern or south-eastern winds in Northern Europe. However, so far the probability for the pest being associated with this pathway is considered as low. This is due to the fact that the following two unusual situations must coincide: a relatively high population density of *L. trifolii* must have been established in the field in a nearby country, and the weather conditions in the area must favor a spread of the population to Norway. This situation could change if *L. trifolii* is established in greenhouses in other countries in Northern Europe.

Because of the long distances from most parts of the world to Norway, pathway C is only possible from Northern Europe.

3.2.1.3 Probability of survival and multiplying during transport or storage

There is a high probability for *L. trifolii* to survive and multiply during transport or storage of host plants (pathway A). This is due to the fact that all the developmental stages of the pest (eggs, larvae, pupae and adult flies) will be able to utilize the host plant for their successful development, and the temperature need of the plants is suitable also for all stages of the pest. The level of uncertainty in this assessment is low.

There is a low probability for *L. trifolii* to survive and multiply during transport or storage of soil or growing media (pathway B). This is due to the fact that pupae can survive periods of approximately 1-2 weeks in soil, away from their host plants. The level of uncertainty in these assessments is low.

3.2.1.4 Probability of pest surviving existing pest management procedures

The likelihood of the pest to survive existing pest management procedures will vary from very unlikely to very likely depending on the commodity and the phytosanitary measures applied. For all pathways and all geographical origins the ability for the pest to remain undetected will be affected by the method of inspection by the exporting country's NPPO and if required by the Norwegian regulations. Similarly, the likelihood of the pest surviving any phytosanitary measures required by Norwegian legislation will depend on the effectiveness of their application and their efficacy. For each pathway ratings of the probability for survival, and uncertainties of the ratings, are given below. So far the Norwegian authorities are of the opinion that *L. trifolii* does not exist in Norway.

Pathway A. Import of host plants with eggs, larvae or pupae

It is moderately likely that *L. trifolii* will survive existing pest management procedures given by Landbruks- og matdepartementet (2000). The known entries into Norway are presented in Table 2. The pest may be present on plants even if the plants originate from an area in which there is an official statement that *L. trifolii* does not occur. It is also moderately likely that the pest will remain undetected on plants that are inspected and tested prior to export to the PRA

area from greenhouses in areas where the pest occurs. The uncertainties of these assessments are low.

Pathway B. Import of soil/growing media with pupae

If *L. trifolii* is present in soil or growing media it is very unlikely to be detected and there is a high probability to survive existing pest management procedures. The uncertainties of these assessments are low.

Pathway C. Natural spread of adult flies by air

Free movement of insects with wind is impossible to control. Consequently, there is a possibility that *L. trifolii* might be wind-borne into the country if the species has an outbreak in a nearby country, or if the species in the future is established in greenhouses or the field in nearby countries. The uncertainty of this assessment is low.

3.2.1.5. Probability of transfer to a suitable host

Due to the polyphagy of *L. trifolii*, the probability of transfer to a suitable host after arrival in the PRA area is high, whatever the way of entry. Regarding the pathway of host plants, the pest is already present on a suitable host. It is very likely that the pest would be transferred to other hosts in Norwegian greenhouses and garden centres. Conditions in greenhouses and garden centers with close spacing of plants favour the dispersal of the pest. Furthermore, *L. trifolii* is very likely to transfer to a suitable environment, when sold to the consumer. The environments of parks and private gardens, at least along the coast of Norway, are very likely to support the pest during summer.

It is highly likely that *L. trifolii* could be transferred from plants in greenhouses to host plants in natural environments during the summer.

3.2.1.6. Summarised probability of entry for each pathway

Pathway A. Import of host plants with eggs, larvae or pupae

The likelihood of *L. trifolii* to enter the PRA area by host plants is rated as high, with low level of uncertainty. This pathway is rated as the most likely pathway for entry of *L. trifolii* into the PRA area.

Pathway B. Import of soil/growing media with pupae

The likelihood of *L. trifolii* to enter the PRA area with contaminated soil is rated as low, with a high level of uncertainty.

Pathway C. Natural spread of adult flies from other European countries by air.

Natural spread of *L. trifolii* by aerial dissemination of adult flies is possible, but the probability is rated as low, with a high level of uncertainty. However, if the species becomes established in nearby countries like Germany, Poland, Denmark or Sweden in the future, such entries will be much more probable.

3.2.2 Probability of establishment

The probability of establishment of *L. trifolii* in the PRA area will vary with the availability of suitable hosts, suitability of the environment, biological characteristics of the pest, and the effects of existing pest management practices. The significance and the uncertainty for each of these topics are addressed in the following paragraphs (4.2.2.1 – 4.2.2.4).

3.2.2.1 Availability of suitable hosts, alternate hosts and vectors in the PRA area

L. trifolii has a very broad host range across a wide range of plant genera and there is an abundant availability of suitable hosts in the PRA area. The uncertainty surrounding this data is low. *Chrysanthemum* sp. has been the imported plant to Norway most commonly infested with *L. trifolii* (Tables 1 and 2). Under natural conditions the pest has infested at least 75 plant species in 59 plant genera worldwide (Appendix 3), representing at least 28 plant families (Table 5). Of these, many grow naturally or in greenhouses in Norway. Wild plants growing in Norway that has been confirmed infested in other countries are among others buttercups (*Ranunculus acris*), although this topic has not been systematically investigated.

Table 5. Plant families that contain host species for *L. trifolii* (Spencer 1990, Sæthre 1996, Andersen *et al.* 2008, EPPO reporting service 2002 – September 2009, EPPO databases on quarantine pests)

Scientific name	Scientific name	Scientific name	Scientific name
<i>Asteraceae</i>	<i>Convolvulaceae</i>	<i>Plantaginaceae</i>	<i>Turneraceae</i>
<i>Alliaceae</i>	<i>Cucurbitaceae</i>	<i>Poaceae</i>	<i>Typhaceae</i>
<i>Alstroemeriaceae</i>	<i>Euphorbiaceae</i>	<i>Primulaceae</i>	<i>Verbenaceae</i>
<i>Amaranthaceae</i>	<i>Fabaceae</i>	<i>Ranunculaceae</i>	<i>Zygophyllaceae</i>
<i>Apiaceae</i>	<i>Iridaceae</i>	<i>Rosaceae</i>	
<i>Brassicaceae</i>	<i>Lamiaceae</i>	<i>Scrophulariaceae</i>	
<i>Caryophyllaceae</i>	<i>Malvaceae</i>	<i>Solanaceae</i>	
<i>Chenopodiaceae</i>	<i>Onagraceae</i>	<i>Tropaeolaceae</i>	

3.2.2.2 Suitability of environment

The environmental conditions in greenhouses in the PRA area are considered to be suitable for *L. trifolii* all year round, with a low level of uncertainty. Outdoors, the environmental conditions are considered to be suitable for *L. trifolii* during the summer in some parts of the PRA area, with a low level of uncertainty. The assessments behind these conclusions are given below.

Climate is an important factor that affects establishment of *L. trifolii*, and climate suitability of the PRA area is therefore analysed in this section. The global distribution of the pest according to EPPO is shown in Appendix 1.

The monthly mean temperature in most parts of coastal Southern Norway in October – May (exemplified by Særheim, Ås and Kvithamar, Table 3) is lower than the lowest developmental temperature of *L. trifolii* of about 10 °C (Lanzoni *et al.* 2002, Tokumaru & Abe 2003).

Consequently, *L. trifolii* could develop in the field only during four months each summer, but would have to stay in the pupal stage for the remaining eight months each winter.

The present distributions show that the polyphagous quarantine *Liriomyza* species cannot successfully overwinter under natural conditions in the temperate areas. However, *Liriomyza* species have dispersed far beyond their apparent overwintering range limit, and in much higher-latitude regions with severe winter conditions, by opportunistic exploitation of protected microhabitats (Kang *et al.* 2009). The climatic conditions necessary for the development of *L. trifolii* has been investigated mainly in Asia during the quick spread of the species towards the north in recent years, and is reported below.

L. trifolii is present in many countries in Asia (Abe & Kawahara 2001, Lee *et al.* 2005, Feng *et al.* 2007, Tokumaru *et al.* 2007, Andersen *et al.* 2008). From the available literature it is difficult to conclude how far north the species could survive the winter outside greenhouses. However, since *L. trifolii* has temperature preferences relatively similar to *L. sativae* (Tokumaru & Abe 2003), it would be reasonable to assume that *L. trifolii* can overwinter in the field north to a latitude of approximately 34° N, with an isotherm of minus 2 °C in January (Zhao & Kang 2000). North of 34° N the species would have to overwinter in greenhouses and infest the fields each year.

L. trifolii was found in the field in agricultural areas in Northern Italy during the summer (Masetti *et al.* 2004). In Portugal it has been established as a pest species, and due to the use of so-called open Mediterranean greenhouses, *L. trifolii* can easily switch between greenhouses and the open field according to preferred temperatures at different times of the year (Godinho & Mexia 2000). These conditions are expected to be similar also in other European countries along the Mediterranean, for instance Spain. Also in Turkey the species has developed into an important pest species in field crops since 1995 (Gencsoylu 2003, Cikman & Cömlekcioglu 2006).

The lower threshold temperature for development of the different larval stages and the pupal stage in different *L. trifolii* populations were found to be 8.0 – 10.1 °C by Saito *et al.* (1995), 9.9 – 10.7 °C by Lanzoni *et al.* (2002), and 8.5 – 11.7 °C by Sakamaki *et al.* (2003). Taking into account the mean temperatures in coastal Norway in the period October – May (Table 5), this means that there will be almost no development of the species in the field during these eight months.

In conclusion, all scientific data suggest that *L. trifolii* will not be able to overwinter in the field even in the mildest areas in Norway. However, when growing host plants continuously in greenhouses, the species will be able to develop large populations. The number of generations will vary with the temperature. At 20 °C the life-cycle takes 23.6 – 25.6 days, and 15 generations would develop per year, while at 25 °C the life-cycle is 15.0 – 16.5 days (Lanzoni *et al.* 2002, Sakamaki *et al.* 2003, Tokumaru & Abe 2003), and 24 generations would develop per year.

3.2.2.3 Cultural practices and control measures

After establishment in greenhouses in the PRA area, the pest will be sought eradicated. In all situations so far this policy has been successful, so it is unlikely that the pest will be established in greenhouses over long periods of time.

Also the managed environment outside greenhouses in parts of the PRA area is favourable for spread of *L. trifolii* during the summer months. It is unlikely that existing pest management practice in the PRA area will prevent spread of the pest in greenhouses or in the field. *L. trifolii* also has many host plants among commonly grown vegetables in Norway. Thus, if

infested greenhouses are in the vicinity of agricultural fields, *L. trifolii* could be spread in fields. However, so far such a situation has not been reported. Based on biological characteristics, it is likely that the pest during summer could survive pest management practices in the field in Norway. The uncertainty surrounding these questions is low.

Likelihood of the existing pest control management practice to prevent establishment of the pest in greenhouses

In Norway, dimethoate, thiacloprid, abamectin, spinosad and several pyrethroids are recommended pesticides against *L. trifolii* and other leafmining flies (Mattilsynet 2009). In addition, two parasitic wasp species and one nematode species are on the current list of biological agents against leafmining flies in Norway. Since eradication has been the chosen strategy by all incidents of *L. trifolii* into the PRA area, the effectiveness of these pest control methods has not been tested. However, due to the experience from control programs in other countries (e.g. Ulubilir & Sekeroglu 1997, Kaspi & Parella 2003, Cikman & Cömlekcioglu 2006), we find it unlikely, with low uncertainty, that these pest management practises currently available in greenhouses, garden centres, parks, private gardens and fields in the PRA area would prevent establishment of *L. trifolii*.

Likelihood to survive eradication programs in the PRA area, based on the biological characteristics of the pest.

So far, eradication has been the chosen strategy by all incidents of *L. trifolii* into the PRA area. Important parts of the eradication program have been full sanitation of infested greenhouses (immediate destruction of all plant material, and heat treatment of the soil) and no growing of potential host plant species for a certain period of time. Due to the successful eradications of *L. trifolii* after all incidents in the PRA area, we find it unlikely that the pest could survive eradication programs in greenhouses in the PRA area. The uncertainty is low. In the field it is highly unlikely that *L. trifolii* can be eradicated by any means during the summer, but during the following winter it will die out.

Suitability of the managed environment in the PRA area for pest establishment

The managed environment around Norwegian greenhouses, garden centres, private gardens and public greens are all favourable to establishment of *L. trifolii* during summer. The uncertainty is low. In greenhouses and garden centres, host plants are abundantly available. Trade networks, which are common between Norwegian greenhouses and garden centres, favour a wider establishment of the pest. In parks, private gardens and natural areas, the environment is also considered favourable due to availability of hosts and conducive climate. Mutual use of equipments at different sites, are examples of management practises that will support the spread and establishment of *L. trifolii*. Once entered into the environment, the pest is favoured by the short generation time and the ready availability of host plants.

3.2.2.4 Other characteristics of the pest affecting the probability of establishment

It is likely that the reproductive strategy of the pest and duration of its life cycle could aid establishment, and it is likely that a population could be spread in the field during the summer months. The pest is highly adaptable and has been introduced into many new areas outside its area of origin. In parts of the temperate zone *L. trifolii* infests crops in the field in summer

even if it cannot overwinter outside greenhouses. This is due to repeated colonization from infested greenhouses every spring. The uncertainty is low for these assessments.

Probability of the reproductive strategy of the pest and the duration of its life cycle to aid establishment.

L. trifolii has a reproduction strategy that most likely would favour a quick spread into the field during summer, and all year round in greenhouses. In greenhouses, at 20 °C the life-cycle takes 23.6 – 25.6 days, and 15 generations would develop per year, while at 25 °C the life-cycle is 15.0 – 16.5 days (Lanzoni *et al.* 2002, Sakamaki *et al.* 2003, Tokumaru & Abe 2003), and 24 generations would develop per year. This rapid development of successive generations is part of the explanation for the quick build-up of huge population. Another important factor both in the greenhouses and in the field is its wide host plant range that makes it probable for it to find host plants everywhere and the development of resistance to many insecticides in many populations. The uncertainty of this assessment is low.

3.2.3 Probability of spread after establishment

There is a high probability for *L. trifolii* to be spread quickly in the PRA area by trade of host plants. The uncertainty of this assessment is low. Planting of infested plants will bring the pest from the greenhouses into the environment.

Spread by natural means

L. trifolii has the opportunity for natural spread in the PRA area during the summer, and it is highly likely that this spread would be rapid. It is assumed that *L. trifolii* in this respect could act similar to *L. huidobrensis*, which in the field in Norway can spread in a circle with diameter at least 1 km during the three summer months (Andersen & Hofsvang 2010).

Long-distance dispersal by natural means includes movement by aerial dissemination of adult flies during major weather events such as wind driven rain and turbulent air. So far this has not been observed in Norway.

Spread by human assistance

There are very high probabilities for *L. trifolii* to be spread quickly by human-mediated means in the PRA area, most significantly through the commercial movement of infested plants for planting. The uncertainty of this assessment is low.

3.2.4 Conclusion on the probability of introduction and spread

Probability of entry

The overall probability of entry of *L. trifolii* into the PRA area is rated as high, with low level of uncertainty. This assessment is based upon identification of pathways, import volume, the probability of the pest being associated with the pathway at origin, the probability of survival and multiplying during transport or storage and the probability of transfer to a suitable host after arrival.

Probability of establishment

The overall probability of establishment in greenhouses of *L. trifolii* in the PRA area is rated as high, with low level of uncertainty. This assessment is based on an abundant availability of suitable hosts, suitability of the environment, biological characteristics of the pest, and the effects of existing pest management practices.

The overall probability of establishment of *L. trifolii* outdoors in the PRA area is rated as low. This assessment is based on the experience with *L. huidobrensis* in 2002 (Andersen & Hofsvang 2010).

Probability of spread after establishment

The probability for *L. trifolii* to be spread quickly in the PRA area by trade of host plants for planting is rated as high. The uncertainty of this assessment is low. Planting of infested plants will bring the pest from the greenhouses into the environment. This can also happen if adult flies escape from the greenhouses through doors or windows. In parts of the PRA area where climate events are favourable, and where there is an abundance of continuous hosts, natural spread is likely to be high during the summer months. During the winter, all out-door populations will die out in the PRA area.

3.3. Assessment of potential economic consequences

3.3.1 Pest effects

3.3.1.1 Direct pest effects

The direct effects by *L. trifolii* include both the biological and aesthetical injury to the plants, cf. 4.1.5. Since *L. trifolii* is listed as a quarantine pest in Norway, all specimens observed have been immediately eradicated. Large infestations have never been observed in Norwegian greenhouses.

3.3.1.2 Indirect pest effects

During the years 1980-1982 three incidents of *L. trifolii* were observed in Norwegian greenhouses (cf. table 2). Some notes from the archives of the Norwegian Plant Protection Institute give some information on the procedures. In all cases destruction of the infested plants in the greenhouses were recommended by the Norwegian Plant Protection Institute, however, but not always performed due to several reasons.

In 1980 one greenhouse with *Chrysanthemum* plants imported from the Canary Islands were infested. *Liriomyza trifolii* was then not included in the list of quarantine pests in Norway, and no order of destruction of the plant material could be issued. Later the same year another greenhouse with *Gerbera* plants imported from the Netherlands was infested. A control program with use different insecticides was launched, and during spring 1981 the greenhouse was declared free of *L. trifolii*, so no destruction of plants proved necessary.

In 1982 another greenhouse with *Chrysanthemum* plants from the Canary Islands was infested with *L. trifolii*. Destruction of 90.000 *Chrysanthemum* cuttings was ordered. This could now be done as *L. trifolii* was added to the Norwegian list of quarantine pests (then named the “A-list”) by Ministry of Agriculture 16 January 1981. The cost of the destructed plants was not evaluated.

During the widespread infestation of *L. huidobrensis* in Norwegian greenhouses and garden centres in 2002 Landbrukstilsynet demanded destruction of all infested plant materials and banned introduction of new plants until the greenhouse was declared free from the quarantine pest (Willumsen 2002). Such actions can have serious economical consequences for the growers involved. The spreading and infestation of *L. huidobrensis* in 2002 was estimated to cost 40-50 million NOK (Miljøverndepartementet 2007).

3.3.2 Analysis of economic consequences

3.3.2.1 Analysis of commercial consequences

The leafminer *L. trifolii* is likely to have significant economic impact in Norwegian greenhouses without current phytosanitary measures. The present regulations with *L. trifolii* as a quarantine pest will initiate immediately destruction of plant material within parts or the complete greenhouse area. The economical consequences will depend on the amount and the total value of plant material destructed in each case.

The level of uncertainty of this assessment is low.

3.3.2.2 Non-commercial and environmental consequences

Appearance of *L. trifolii* in natural areas in the PRA area could be a local threat to closely related species, mainly *L. bryoniae* and *L. strigata*, by competing over host plants. In addition it would cause locally high infestations in host plants. However, since the species will be eradicated during the winter, the threat to the environment, both plants and animals, is valuated as low.

3.3.3 Conclusion of the assessment of economic consequences

3.3.3.1 Endangered area

The PRA area where presence of *L. trifolii* might result in economically important losses is identified as Norwegian greenhouses and garden centres.

The pest *L. trifolii* is likely to have moderate economic impact on the garden centres and greenhouses in the PRA area with current phytosanitary measures. Without any such regulations *L. trifolii* would likely have major economic impact in greenhouses in the PRA area.

L. trifolii is likely to have a low economic impact on outdoor crops the PRA area.

4. CONCLUSION OF THE PEST RISK ASSESSMENT

Pest status of the PRA area

The pest of concern in this pest risk assessment is the Agromyzid fly *Liriomyza trifolii*. The PRA area is Norway. *L. trifolii* is not present, and the pest is a quarantine species in the PRA area. It has been imported several times, but each time it has been eradicated.

Probability of introduction and spread

The overall probability of entry of *L. trifolii* into the PRA area is rated as high. This assessment is based upon identification of pathways, import volume, the probability of the pest being associated with the pathway at origin, the probability of survival and multiplying during transport or storage and the probability of transfer to a suitable host after arrival.

The overall probability of establishment of *L. trifolii* in greenhouses in the PRA area is rated as high. The probability of establishment in the field is rated as high during the summer months, but its ability to overwinter in the field in the PRA area is evaluated as very low. This assessment is based on an abundant availability of suitable hosts, suitability of the environment in at least parts of the PRA area, and biological characteristics of the pest. The uncertainty of these assessments is low.

Conclusion regarding endangered areas

The part of the PRA area where presence of *L. trifolii* in greenhouses might result in economically important losses (the endangered area) is assessed to be all of the country of Norway. This area must be regarded as a maximum estimate for the endangered area. In the field, the species would need a summer temperature of at least 15 °C to develop populations of a certain size to become a pest. This could happen in coastal areas of Southern and Middle Norway.

Conclusion of the assessment of economic consequences

The pest *L. trifolii* is likely to have moderate economic impact on the garden centres and greenhouses in the PRA area with current phytosanitary measures. Without any such regulations *L. trifolii* would likely have major economic impact in greenhouses in the PRA area.

L. trifolii is likely to have a low economic impact on outdoor crops the PRA area. The uncertainty of these assessments is low.

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Appendix 1

Appendix 1. Global distribution of *Liriomyza trifolii* (EPPO 2009)

Country	Widespread	Limited distribution	Few records	No details
Europe				
Cyprus	X			
France	X			
Italy	X			
Spain	X			
Austria		X		
Belgium		X		
Croatia		X		
Greece		X		
Netherlands		X		
Portugal		X		
Russia		X		
Slovenia		X		
Turkey		X		
Norway			X	
Switzerland			X	
Malta				X
Romania				X
Asia				
Israel	X			
Taiwan			X	
China				X
India				X
Japan				X
Jordan				X
Korean republic				X
Lebanon				X

Oman				X
Phillippines				X
Saudi Arabia				X
Yemen				X
Africa				
Senegal	X			
Tunisia		X		
South Africa			X	
Benin				X
Cote d'Ivoire				X
Egypt				X
Ethiopia				X
Guinea				X
Kenya				X
Madagascar				X
Mauritius				X
Mayotte				X
Morocco				X
Nigeria				X
Reunion				X
Sudan				X
Tanzania				X
Zambia				X
Zimbabwe				X
America				
Martinique	X			
Canada		X		
Chile		X		
Netherlands Antilles		X		
USA		X		
Argentina				X
Bahamas				X

Barbados				X
Bermuda				X
Brazil				X
Colombia				X
Costa Rica				X
Cuba				X
Dominican Republic				X
Ecuador				X
French Guiana				X
Gualadeloupe				X
Guatemala				X
Guyana				X
Mexico				X
Peru				X
Puerto Rico				X
Trinidad and Tobago				X
Venezuela				X
Virgin Islands (British)				X
Virgin Islands (USA)				X
Oceania				
American Samoa				X
Guam				X
Micronesia				X
Northern Mariana Islands				X
Samoa				X
Tonga				X

Appendix 2

Appendix 2. EPPO report on notifications of non-compliance for *Liriomyza trifolii* (records of *Liriomyza* spp. are not included) (EPPO Reporting Service 2002 – September 2009).

Year	Consignment	Type of commodity	Country of origin	Destination
2009	<i>Apium graveolens</i>	Vegetables	Thailand	Netherlands
	<i>Apium graveolens</i>	Vegetables	Thailand	Sweden
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Ocimum</i>	Vegetables (leaves)	Thailand	Sweden
	<i>Ocimum americanum</i>	Vegetables (leaves)	Thailand	France
	<i>Ocimum americanum</i>	Vegetables (leaves)	Thailand	Sweden
	<i>Ocimum basilicum</i>	Vegetables (leaves)	Thailand	Sweden
	<i>Apium graveolens</i>	Vegetables	Thailand	Netherlands
	<i>Apium graveolens</i>	Vegetables	Thailand	Sweden
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Ocimum</i>	Vegetables (leaves)	Thailand	Sweden
	<i>Ocimum americanum</i>	Vegetables (leaves)	Thailand	France
	<i>Ocimum americanum</i>	Vegetables (leaves)	Thailand	Sweden
	<i>Ocimum basilicum</i>	Vegetables (leaves)	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Ocimum americanum</i>	Vegetables (leaves)	Thailand	Sweden
	<i>Apium graveolens</i>	Vegetables	Thailand	Netherlands
	<i>Apium graveolens</i>	Vegetables	Thailand	Sweden
	<i>Ocimum americanum</i>	Vegetables (leaves)	Thailand	Sweden
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
2008	<i>Ocimum basilicum</i>	Vegetables (leaves)	Thailand	Denmark
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Apium graveolens</i>	Vegetables	Thailand	Denmark
	<i>Apium graveolens</i>	Vegetables	Thailand	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands

	<i>Gypsophila</i>	Cut flowers	Kenya	Netherlands
	<i>Solidago</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Aster</i>	Cut flowers	Israel	Netherlands
	<i>Chrysanthemum</i>	Cut flowers	Colombia	Netherlands
	<i>Chrysanthemum</i>	Cut flowers	Costa Rica	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Apium graveolens</i>	Vegetables	Thailand	Denmark
	<i>Chrysanthemum</i>	Cut flowers	Costa Rica	Netherlands
	<i>Gypsophila</i>	Cut flowers	Ethiopia	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
2007	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Aster</i>	Cut flowers	Israel	Netherlands
	<i>Chrysanthemum</i>	Cut flowers	Colombia	Netherlands
	<i>Eustoma</i>	Cut flowers	Turkey	Netherlands
	<i>Gypsophila</i>	Cut flowers	Egypt	Netherlands
	<i>Gypsophila</i>	Cut flowers	Ethiopia	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Aster, Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Chrysanthemum</i>	Cut flowers	Netherlands	Russia
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Chrysanthemum</i>	Cut flowers	South Africa	Netherlands
	<i>Eustoma</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Ethiopia	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	United Kingdom
	<i>Ocimum basilicum</i>	Vegetables (leaves)	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
2006	<i>Eustoma</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Ethiopia	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands

	<i>Gypsophila</i>	Cut flowers	Israel	United Kingdom
	<i>Ocimum basilicum</i>	Vegetables	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila, Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Trachelium</i>	Cut flowers	Israel	Netherlands
	<i>Aster</i>	Cut flowers	Israel	Netherlands
	<i>Eustoma</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Egypt	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila, Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Trachelium</i>	Cut flowers	Israel	Netherlands
2005	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Lisianthus</i>	Cut flowers	Israel	Netherlands
	<i>Lisianthus, Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila paniculata</i>	Cut flowers	Israel	United Kingdom
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Solidago</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Tanzania	Netherlands
	<i>Dendranthema</i>	Cut flowers	Spain (Canary isl.)	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Ranunculus, Asclepias, Eustoma</i>	Cut flowers	Israel	Netherlands
2004	<i>Aster</i>	Cut flowers	Costa Rica	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands

	<i>Gypsophila</i>	Cut flowers	Tanzania	Netherlands
	<i>Aster, Solidago</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Solidago</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Gypsophila paniculata</i>	Cut flowers	Spain	United Kingdom
2003	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Solidago hybrida</i>	Cut flowers	Zimbabwe	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila, Rosa</i>	Cut flowers	Israel	Netherlands
	<i>Gypsophila</i>	Cut flowers	Israel	Netherlands
	<i>Chrysanthemum</i>	Cut flowers	Netherlands	United Kingdom
	<i>Gypsophila</i>	Cut flowers	Israel	United Kingdom
	<i>Ocimum basilicum</i>	Vegetables	Israel	France
	<i>Gerbera</i>	Plants for planting	Netherlands	Finland
	<i>Gerbera</i>	Cuttings	USA	Netherlands
2002	<i>Dendranthema</i>	Cut flowers	Netherlands	United Kingdom
	<i>Gypsophila</i>	Plants for planting	Netherlands	Lithuania
	<i>Gypsophila paniculata</i>	Cut flowers	Italy	United Kingdom
	<i>Echinacea</i>	Cuttings	Netherlands	United Kingdom

Appendix 3

Appendix 3. Host plants for *Liriomyza trifolii* (Sæthre 1996, Andersen *et al.* 2008, EPPO reporting service 2002 – September 2009, EPPO databases on quarantine pests).

Host species	Major host = A; Minor host or not classified = B
<i>Allium cepa</i>	B
<i>Allium porrum</i>	B
<i>Allium sativum</i>	B
<i>Alstromeria</i> sp.	B
<i>Anethum graveolens</i>	B
<i>Anthriscus cereifolium</i>	B
<i>Antirrhinum</i> sp.	B
<i>Apium graveolens</i>	A
<i>Apium petroselinum</i>	B
<i>Arachis hypogaea</i>	B
<i>Aster novi-belgii</i>	B
<i>Aster</i> sp.	B
<i>Beta vulgaris</i>	B
<i>Bidens pilosa</i>	B
<i>Bidens</i> sp.	B
<i>Brassica sinensis</i>	B
<i>Brassica juncea</i>	A
<i>Brassica oleracea</i>	B
<i>Calendula officinalis</i>	B
<i>Capsella bursa-pastoris</i>	B
<i>Capsicum annuum</i>	B
<i>Chenopodium album</i>	B
<i>Chrysanthemum frutescens</i>	B
<i>Chrysanthemum morifolium</i>	A
<i>Chrysanthemum</i> sp.	B
<i>Citrullus lanatus</i>	B
<i>Cucumis melo</i>	B
<i>Cucumis sativus</i>	B
<i>Cucurbita pepo</i>	B

<i>Dahlia</i> sp.	B
<i>Dahlia</i> hybrids	B
<i>Dendranthema indicum</i>	B
<i>Dendranthema</i> x <i>grandiflorum</i>	A
<i>Dendranthema</i> sp.	B
<i>Dianthus caryophyllus</i>	B
<i>Dianthus</i> sp.	B
<i>Echinacea</i> sp.	B
<i>Epilobium</i> sp.	B
<i>Eustoma</i> sp.	B
<i>Gerbera jamesonii</i>	B
<i>Gerbera</i> sp.	B
<i>Gossypium hirsutum</i>	B
<i>Gypsophila paniculata</i>	B
<i>Gypsophila</i> sp.	B
<i>Helianthus bispinatus</i>	B
<i>Hibiscus esculentus</i>	B
<i>Lactuca sativa</i>	B
<i>Lathyrus</i> sp.	B
<i>Leucanthemum vulgare</i>	B
<i>Leucanthemum</i> x <i>superbum</i>	B
<i>Lycopersicon esculentum</i>	B
<i>Medicago sativa</i>	B
<i>Ocimum americanum</i>	B
<i>Ocimum basilicum</i>	B
<i>Ocimum</i> sp.	B
<i>Pericallis</i> x <i>hybrida</i>	B
<i>Phaseolus coccineus</i>	B
<i>Phaseolus lunatus</i>	B
<i>Phaseolus vulgaris</i>	B
<i>Pisum sativum</i>	B
<i>Plantago lanceolata</i>	B
<i>Plantago major</i>	B
<i>Ranunculus repens</i>	B

<i>Ranunculus</i> sp.	B
<i>Raphanus sativus</i>	B
<i>Senecio cruentus</i>	B
<i>Senecio jacobaea</i>	B
<i>Senecio vulgaris</i>	B
<i>Solanum dulcamara</i>	B
<i>Solanum tuberosum</i>	B
<i>Solidago hybrida</i>	B
<i>Solidago</i> sp.	B
<i>Spilanthus acmello</i>	B
<i>Spinacia oleraceae</i>	B
<i>Tagetes</i> sp.	B
<i>Tanacetum parthenium</i>	B
<i>Tanacetum vulgare</i>	B
<i>Trachelium</i> sp.	B
<i>Trifolium</i> sp	B
<i>Tropaeolum majus</i>	B
<i>Tropaeolum</i> sp.	B
<i>Verbena</i> sp	B
<i>Vicia faba</i>	B
<i>Vigna</i> sp.	B
<i>Zinnia</i> sp.	B